

Sudden Oak Death

Yvonne Barkley



Sudden oak death syndrome (SOD) was first reported in the Netherlands and Germany in 1993, causing leaf-blight, stem cankers, and tip dieback on nursery grown rhododen-

drons and viburnums. SOD has become the latest serious threat to plant materials across the United States and was first spotted in the U.S. in 1995 on tanoak in a Marin County, CA forest. It took University of California researchers until 2001 to positively identify the pathogen, after which the USDA imposed a federal quarantine of 12 California counties.

During the summer of 2001, SOD was also found killing wild rhododendrons in Brookings, OR. Infected plants were eradicated soon after sites were found and this infestation remains confined within a 9-mile quarantined area of forested land near Brookings. The initial source of infection of these sites remains unknown.

In the spring of 2003, SOD was found in a wholesale nursery in Portland, OR and also in a retail outlet in Washington that received plants from the Oregon wholesaler. Genera affected were Pieris, Viburnum, and Rhododendron. Infected stock and other nearby host plants were destroyed. This outbreak is suspected to have come from an international shipment of rhododendrons.

On March 25, 2004, SOD was confirmed on plants at Monrovia Growers in Azusa, CA and Specialty Plants, Inc. in San Marcos, CA. On March 29, 2004, the US Department of Agriculture put all 1,500 nurseries in California under a 60-day quarantine. Each nursery has been inspected for symptoms of SOD and released if no symptoms were found.

To date, there are confirmed trace forwards on stock from Monrovia in 14 states: California, Alabama, Florida, Washington, Oregon, Texas, Colorado, Georgia, Louisiana, Maryland, North Carolina, New Mexico, Tennessee, and Virginia (Situation Report #43, May 25, 2004).

Cause and Hosts

SOD is caused by the fungus-like organism Phytophthora ramorum (P. ramorum), and causes different symptoms on different hosts. Called sudden oak death when it occurs on susceptible tree species, P. ramorum is characterized by bleeding cankers that girdle the trunks. On affected shrub species, the disease is called Ramorum leaf and shoot blight and is characterized by leaf blights and shoot diebacks. Unfortunately, on many plant species symptoms of P. ramorum may be indistinguishable from other common Phytophthora fungal diseases.

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Phytophthora ramorum is well adapted to cool, wet conditions and tolerates heat and drought. Unlike other species in this large fungal family that usually infect roots, P. ramorum is mainly a foliar pathogen. It produces several types of spores, which helps the organism survive and spread. Spores landing on wet leaves or stems germinate and infect young plant tissues. Several of the spore types can be moved with windborne rain, in irrigation water, or with water splashed onto foliage by sprinkler irrigation. P. ramorum is also thought to move in soil and on infected plant material. As of this writing there are 60 hosts and associated host species in at least 12 plant families (see Table 1 above and Table 2 on page 4 and 5).

Symptoms

On trees. P. ramorum can affect just the bark, as on beech, or both the bark and the leaves, as on tanoak. It is also possible that some trees like ash, which has highly susceptible leaves experimentally (but has not yet been found to be a natural host) may just be leaf hosts to the pathogen.



Bark infections occur as large cankers on the lower trunk that are brown to black discolored areas on the outer bark and "bleed" dark red sap. Mottled areas of dead and discolored inner bark with black lines around the edges are visible when the bark is

removed. A rapid change in the color and condition of the foliage, followed by sudden death of the tree, occur when cankers girdle the trunks of affected trees. Cankers can cause sudden death, as in California tanoak, or a slow decline over several years, as in American oak species. Cankers do not extend below the soil line and do not appear to infect the roots.



On leaves. Leaf infections appear as brown to black dead areas, often at the tip or base of a leaf. Shoots can also be affected and appear brown to black in color and may wilt.

Symptoms on specific species include:

- On conifers in the U.S.: needle blight and dieback of young shoots of Douglas fir, coastal redwood, and grand fir.
- On yew: needle blight of young foliage resulting in aerial dieback.
- On Rhododendron, Pieris, Vaccinium, and Hamamelis spp. (witchhazel): affected shoots or twigs develop a brown to black discoloration that spreads along the twig and can spread into the leaf via the petiole; characteristics symptoms are a blackening of the petiole, leaf base, and leaf tip that may extend along the mid-vein; twig cankers lead to wilting of affected shoots; development of symptoms can be rapid.
- On viburnum: infection commonly occurs at the base of the stem, causing wilting and then death. May also cause brown to black leaf infections and can also affect flowers, especially on evergreen species.
- On Camellia, Kalmia, Syringa (lilac) and Leucothoe spp.: usually confined to leaf infections; leaf lesions are brown to black and typically occur at the tip or edges of the leaf; some shoots may become infected, leading to dieback.

Management

Management efforts have been focused on eradicating the pathogen where it is found and preventing new infections. Early detection is vital to preventing the disease from spreading. In order to limit the spread of this disease, homeowners with a suspected plant infected with P. ramorum should contact your nearest Extension Office and ask them about collecting a sample of the plant.

Growers may submit a sample directly. PLEASE FOLLOW THE PROTOCOL LISTED BELOW.

1. Suspect plant material must be placed in double ziplock bags and stored in a refrigerator awaiting shipment to a diagnostic facility. The preferred method for

shipment is triple packaging: two ziplock bags and an outer container. The outer shipping container should be an approved cardboard shipping box. The seams of the box should be closed with approved shipping tape.

2. Include the following information with the sample:

- Host.
- Collector's name, and date.
- Number of hosts at that site.
- Facility/location name and address, contact person, and county.
- Other potential hosts at the site and any observations.

3. Samples should be shipped via overnight delivery or hand delivered to:

S. Krishna Mohan Professor of Plant Pathology University of Idaho 29603 U of I Lane Parma, ID 83660 Phone: (208) 722-6701 Ext. 218 Fax: (208) 722-6708

4. All tools and other equipment must be sanitized and/or sterilized before re-use.

Control

Phytophthora ramorum has only recently been described and knowledge of how the disease develops and spreads is limited. Currently, fungicides are being used as preventative treatments on known host plants. Most of the fungicides used to control other Phytophthora species do not control this organism; they can only prevent it from becoming established.

For more information on Phytophthora ramorum go to the USDA APHIS website at:

www.aphis.usda.gov/ppq/ispm/sod/

Parts of this article were excerpted from:

Feiber, D. 2004. Sudden Oak Death Detected on Plants Imported into Florida. News Release, Florida Department of Agriculture and Consumer Services, Tallahassee, FL.

Parke, J. J. Pscheidt, and R. Linderman. 2003. Phytophthora ramorum: a guide for Oregon nurseries. EM 8840, Oregon State University, Extension Service. 8 pp.

Unknown. 2001. Phytophthora ramorum (Sudden Oak Death). Pest Fact Sheet, Plant Health Risk Assessment Unit, Science Division, Canadian Food Inspection Agency, Pacific Forestry Centre, Canadian Forest Service, Natural Resources, Canada.

Unknown. 2004. Phytophthora ramorum: a threat to our trees, woodland and heartland. DEFRA, Edinburgh, UK. www.defra.gov.uk/planth/ph.htm

Unknown. 2004. USDA Announces Oak Disease Detection Program for California Nurseries. News Release, USDA APHIS Program Update, March 29, 2004.

Unknown. 2004. New Restrictions for P. ramorum. News Release, USDA APHIS Program Update, April 9, 2004.

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Table 1. Plant species currently regulated for Phytophthora ramorum^{*} (in whole or part) – June 1, 2004 – see www.aphis.usda.gov/ppq/ispm/sod

Scientific Name	Common Name
Acer macrophyllum	bigleaf maple
Aesculus californica	California buckeye
Arbutus menziesii	madrone
Arctostaphylos manzanita	manzanita
Camellia japonica	Japanese camellia
Camellia sasanqua	Sasanqua camellia
Hamamelis virginiana	witch hazel
Heteromeles arbutifolia	toyon
Lithocarpus densiflorus	tanoak
Lonicera hispidula	California honeysuckle
Pieris formosa	Himalaya pieris
Pieris formosa x japonica	Pieris 'Forest Flame'
Pieris floribunda x japonica	Pieris 'Brouwer's Beauty'
Pieris japonica	Japanese pieris
Pseudotsuga menziesii var. menziesii	Douglas-fir
Quercus agrifolia	coast live oak
Quercus chrysolepis	canyon live oak
Quercus kelloggii	California black oak
Quercus parvula v. shrevei	Shreve's oak
Rhamnus californica	California coffeeberry
Rhododendron spp.	rhododendron (including azalea)
Rosa gymnocarpa	Wood's rose
Sequoia sempervirens	coast redwood
Trientalis latifolia	western starflower
Umbellularia californica	California bay laurel, Oregon myrtle
Vaccinium ovatum	evergreen huckleberry
Viburnum x bodnantense	bodnant Viburnum
Viburnum plicatum var. tomentosum	doublefile Viburnum
Viburnum tinus	laurustinus

* Regulated plants are those adapted from other regulated lists or were added upon completion, documentation and review of traditional Kock's postulates. Some are regulated in part (such as redwood and Douglas-fir); others are regulated in their entirety (such as tanoak and western star flower). Details on regulated articles can be found via links to "Phytophthora ramorum 7 CFR 301.92" and "Recent Modifications to Phytophthora ramorum Regulations" at www.aphis.usda.gov/ppq/ispm/sod

Table 2. Plants Associated with Phytophthora ramorum* (in whole or part) – June 1, 2004 – see www.aphis.usda.gov/ppq/ispm/sod

Common Name

Abies grandis	grand fir
Aesculus hippocastanum	horse-chestnut
Arbutus unedo	strawberry tree
Camellia reticulata	Camellia
Camellia x williamsii	Camellia
Castanea sativa	sweet chestnut
Corylus cornuta	California hazelnut
Fagus sylvatica	European beech
Kalmia latifolia	mountain laurel
Leucothoe fontanesiana	drooping leucothoe
Pieris formosa var. forrestii	Chinese pieris
Pieris formosa var. forrestii x Pieris japonica	Pieris
Pittosporum undulatum	victorian box
Pyracantha koidzumii	Formosa firethorn
Quercus cerris	European turkey oak
Quercus falcata	southern red oak
Quercus ilex	Holm oak
Quercus rubra	northern red oak
Rhamnus purshiana	Cascara
Rubus spectabilis	salmonberry
Syringa vulgaris	lilac
Taxus baccata	European yew
Toxicodendron diversiloba	poison oak
Vaccinium vitis-idaea	lingonberry
Viburnum davidii	David Viburnum
Viburnum farreri (=V. fragrans)	fragrant Viburnum
Viburnum lantana	wayfaringtree Viburnum
Viburnum opulus	European cranberrybush Viburnum
Viburnum x burkwoodii	Burkwood Viburnum
Viburnum x carlcephalum x V. utile	viburnum
Viburnum x pragense	Prague Viburnum

* Plants Associated with Phytophthora ramorum: Associated plants are those found naturally infected from which P. ramorum has been cultured and/or detected using PCR (Polymerase Chain Reaction). For each of these, traditional Koch's postulates have not yet been completed or documented and reviewed.

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